Policy Statement

Unless approved in another policy, genetic cancer susceptibility panel testing (e.g., pan cancer or large panels) is considered investigational.

Multi-gene panel testing for hereditary cancers other than breast, ovarian, colorectal, and non-small-cell lung cancer (see Policy Guidelines) are considered investigational.

Policy Guidelines

Limited Panel Testing
Some limited panel testing may be considered medically necessary when criteria are met as addressed in other Blue Shield of California medical policies specific to those panels.

Although large genetic cancer susceptibility panel testing, is considered investigational, there may be individual or smaller components of the panel that are medically necessary.

Testing related to hereditary breast and ovarian cancer, see Blue Shield of California Medical Policy: Genetic Testing for BRCA1 or BRCA2 for Hereditary Breast/Ovarian Cancer Syndrome and Other High-Risk Cancers.

Testing related to hereditary colorectal cancer, see Blue Shield of California Medical Policy: Genetic Testing for Lynch Syndrome and Other Inherited Colon Cancer Syndromes.

Panel testing related to non-small-cell lung cancer, see Blue Shield of California Medical Policy: Circulating Tumor DNA for Management of Non-Small-Cell Lung Cancer (Liquid Biopsy).

Comprehensive Variant Analysis
Comprehensive variant analysis currently includes sequencing the coding regions and intron and exon splice sites, as well as testing to detect common large deletions and rearrangements that can be missed with sequence analysis alone. In addition, before August 2006, testing for large deletions and rearrangements was not performed, thus some patients with familial breast cancer who had negative BRCA testing before this time may consider repeat testing for the rearrangements (see Policy section for criteria).

Genetics Nomenclature Update
The Human Genome Variation Society nomenclature is used to report information on variants found in DNA and serves as an international standard in DNA diagnostics. It is being implemented for genetic testing medical evidence review updates starting in 2017 (see Table PG1). The Society’s nomenclature is recommended by the Human Variome Project, the HUman Genome Organization, and by the Human Genome Variation Society itself.

The American College of Medical Genetics and Genomics and the Association for Molecular Pathology standards and guidelines for interpretation of sequence variants represent expert opinion from both organizations, in addition to the College of American Pathologists. These recommendations primarily apply to genetic tests used in clinical laboratories, including genotyping, single genes, panels, exomes, and genomes. Table PG2 shows the recommended standard terminology—“pathogenic,” “likely pathogenic,” “uncertain significance,” “likely benign,” and “benign”—to describe variants identified that cause Mendelian disorders.
Table PG1. Nomenclature to Report on Variants Found in DNA

<table>
<thead>
<tr>
<th>Previous</th>
<th>Updated</th>
<th>Definition</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mutation</td>
<td>Disease-associated variant</td>
<td>Disease-associated change in the DNA sequence</td>
</tr>
<tr>
<td></td>
<td>Change in the DNA sequence</td>
<td></td>
</tr>
<tr>
<td>Familial variant</td>
<td>Disease-associated variant identified in a proband for use in subsequent targeted genetic testing in first-degree relatives</td>
<td></td>
</tr>
</tbody>
</table>

Table PG2. ACMG-AMP Standards and Guidelines for Variant Classification

<table>
<thead>
<tr>
<th>Variant Classification</th>
<th>Definition</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pathogenic</td>
<td>Disease-causing change in the DNA sequence</td>
</tr>
<tr>
<td>Likely pathogenic</td>
<td>Likely disease-causing change in the DNA sequence</td>
</tr>
<tr>
<td>Variant of uncertain significance</td>
<td>Change in DNA sequence with uncertain effects on disease</td>
</tr>
<tr>
<td>Likely benign</td>
<td>Likely benign change in the DNA sequence</td>
</tr>
<tr>
<td>Benign</td>
<td>Benign change in the DNA sequence</td>
</tr>
</tbody>
</table>

American College of Medical Genetics and Genomics; AMP: Association for Molecular Pathology.

Genetic Counseling

Experts recommend formal genetic counseling for patients who are at risk for inherited disorders and who wish to undergo genetic testing. Interpreting the results of genetic tests and understanding risk factors can be difficult for some patients; genetic counseling helps individuals understand the impact of genetic testing, including the possible effects the test results could have on the individual or their family members. It should be noted that genetic counseling may alter the utilization of genetic testing substantially and may reduce inappropriate testing; further, genetic counseling should be performed by an individual with experience and expertise in genetic medicine and genetic testing methods.

Coding

There are CPT codes for genomic sequencing procedures (or next-generation sequencing panels). If the panel meets the requirements listed in the code descriptor, the following codes may be used:

- **81432**: Hereditary breast cancer-related disorders (e.g., hereditary breast cancer, hereditary ovarian cancer, hereditary endometrial cancer); genomic sequence analysis panel, must include sequencing of at least 14 genes, including ATM, BRCA1, BRCA2, BRIp1, CDH1, MLH1, MSH2, MSH6, NBN, PALB2, PTEN, RAD51C, STK11, and TP53
- **81433**: Hereditary breast cancer-related disorders (e.g., hereditary breast cancer, hereditary ovarian cancer, hereditary endometrial cancer); duplication/deletion analysis panel, must include analyses for BRCA1, BRCA2, MLH1, MSH2, and STK11
- **81435**: Hereditary colon cancer disorders (e.g., Lynch syndrome, PTEN hamartoma syndrome, Cowden syndrome, familial adenomatosis polyposis); genomic sequence analysis panel, must include sequencing of at least 10 genes, including APC, BMPR1A, CDH1, MLH1, MSH2, MSH6, MUTYH, PTEN, SMAD4, and STK11
- **81436**: Hereditary colon cancer disorders (e.g., Lynch syndrome, PTEN hamartoma syndrome, Cowden syndrome, familial adenomatosis polyposis); duplication/deletion analysis panel, must include analysis of at least 5 genes, including MLH1, MSH2, EPCAM, SMAD4, and STK11
- **81437**: Hereditary neuroendocrine tumor disorders (e.g., medullary thyroid carcinoma, parathyroid carcinoma, malignant pheochromocytoma or paraganglioma); genomic sequence analysis panel, must include sequencing of at least 6 genes, including MAX, SDHB, SDHC, SDHD, TMEM127, and VHL
- **81438**: Hereditary neuroendocrine tumor disorders (e.g., medullary thyroid carcinoma, parathyroid carcinoma, malignant pheochromocytoma or paraganglioma); duplication/deletion analysis panel, must include analyses for SDHB, SDHC, SDHD, and VHL
- **81445**: Targeted genomic sequence analysis panel, solid organ neoplasm, DNA analysis, and RNA analysis when performed, 5-50 genes (e.g., ALK, BRAF, CDKN2A, EGFR, ERBB2,
KIT, KRAS, NRAS, MET, PDGFA, PDGFRB, PGR, PIK3CA, PTEN, RET), interrogation for sequence variants and copy number variants or rearrangements, if performed

- **81450**: Targeted genomic sequence analysis panel, hematolymphoid neoplasm or disorder, DNA analysis, and RNA analysis when performed, 5-50 genes (e.g., BRAF, CEBPA, DNM3A, EZH2, FLT3, IDH1, IDH2, JAK2, KRAS, KIT, MLL, NPM1, NOTCH1), interrogation for sequence variants, and copy number variants or rearrangements, or isoform expression or mRNA expression levels, if performed

- **81455**: Targeted genomic sequence analysis panel, solid organ or hematolymphoid neoplasm, DNA analysis, and RNA analysis when performed, 51 or greater genes (e.g., ALK, BRAF, CDKN2A, CEBPA, DNM3A, EGFR, ERBB2, EZH2, FLT3, IDH1, IDH2, JAK2, KIT, KRAS, MLL, NPM1, NRAS, MET, NOTCH1, PDGFRA, PDGFRB, PGR, PIK3CA, PTEN, RET), interrogation for sequence variants and copy number variants or rearrangements, if performed

**Effective October 1, 2019**, there is a new CPT PLA code that represents BRCAPlus by Ambry Genetics:

- **0129U**: Hereditary breast cancer-related disorders (e.g., hereditary breast cancer, hereditary ovarian cancer, hereditary endometrial cancer), genomic sequence analysis and deletion/duplication analysis panel (ATM, BRCA1, BRCA2, CDH1, CHEK2, PALB2, PTEN, and TP53)

**Effective October 1, 2019**, there is a new CPT PLA code that represents +RNAInsight™ for ColoNext® by Ambry Genetics:

- **0130U**: Hereditary colon cancer disorders (e.g., Lynch syndrome, PTEN hamartoma syndrome, Cowden syndrome, familial adenomatous polyposis), targeted mRNA sequence analysis panel (APC, CDH1, CHEK2, MLH1, MSH2, MSH6, MUTYH, PMS2, PTEN, and TP53) (List separately in addition to code for primary procedure)

**Effective October 1, 2019**, there is a new CPT PLA code that represents +RNAInsight™ for BreastNext® by Ambry Genetics:

- **0131U**: Hereditary breast cancer-related disorders (e.g., hereditary breast cancer, hereditary ovarian cancer, hereditary endometrial cancer), targeted mRNA sequence analysis panel (13 genes) (List separately in addition to code for primary procedure)

**Effective October 1, 2019**, there is a new CPT PLA code that represents +RNAInsight™ for OvaNext® by Ambry Genetics:

- **0132U**: Hereditary ovarian cancer-related disorders (e.g., hereditary breast cancer, hereditary ovarian cancer, hereditary endometrial cancer), targeted mRNA sequence analysis panel (17 genes) (List separately in addition to code for primary procedure)

**Effective October 1, 2019**, there is a new CPT PLA code that represents +RNAInsight™ for ProstateNext® by Ambry Genetics:

- **0133U**: Hereditary prostate cancer-related disorders, targeted mRNA sequence analysis panel (11 genes) (List separately in addition to code for primary procedure)

**Effective October 1, 2019**, there is a new CPT PLA code that represents +RNAInsight™ for CancerNext® by Ambry Genetics:

- **0134U**: Hereditary pan cancer (e.g., hereditary breast and ovarian cancer, hereditary endometrial cancer, hereditary colorectal cancer), targeted mRNA sequence analysis panel (18 genes) (List separately in addition to code for primary procedure)

**Effective October 1, 2019**, there is a new CPT PLA code that represents +RNAInsight™ for GYNPlus® by Ambry Genetics:

- **0135U**: Hereditary gynecological cancer (e.g., hereditary breast and ovarian cancer, hereditary endometrial cancer, hereditary colorectal cancer), targeted mRNA
sequence analysis panel (12 genes) (List separately in addition to code for primary procedure)

**Effective October 1, 2019**, there is a new CPTPLA code that represents +RNAinsight™ for ATM by Ambry Genetics:
- **0136U**: ATM (ataxia telangiectasia mutated) (e.g., ataxia telangiectasia) mRNA sequence analysis (List separately in addition to code for primary procedure)

**Effective October 1, 2019**, there is a new CPTPLA code that represents +RNAinsight™ for PALB2 by Ambry Genetics:
- **0137U**: PALB2 (partner and localizer of BRCA2) (e.g., breast and pancreatic cancer) mRNA sequence analysis (List separately in addition to code for primary procedure)

**Effective October 1, 2019**, there is a new CPTPLA code that represents +RNAinsight™ for BRCA1/2 by Ambry Genetics:
- **0138U**: BRCA1 (BRCA1, DNA repair associated), BRCA2 (BRCA2, DNA repair associated) (e.g., hereditary breast and ovarian cancer) mRNA sequence analysis (List separately in addition to code for primary procedure)

If the panel does not meet the criteria in the specific code descriptors or if the specific analyte is not listed in the more specific CPT codes, then unlisted code 81479 would be reported. The unlisted code would be reported once to represent all of the unlisted analytes in the panel.

### Description

Commercially available cancer susceptibility gene panels can test for multiple variants associated with a specific type of cancer or can include variants associated with a wide variety of cancers. Some of these variants are associated with inherited cancer syndromes. The cancer type(s), as well as a cancer history involving multiple family members, increase the clinical concern for the presence of a heritable genetic variant. It has been proposed that variant testing using next-generation sequencing technology to analyze multiple genes at one time (panel testing) can optimize genetic testing in these patients compared with sequencing single genes.

### Related Policies

- General Approach to Evaluating the Utility of Genetic Panels
- Genetic Testing for BRCA1 or BRCA2 for Hereditary Breast/Ovarian Cancer Syndrome and Other High-Risk Cancers
- Genetic Testing for Li-Fraumeni Syndrome
- Genetic Testing for Lynch Syndrome and Other Inherited Colon Cancer Syndromes
- Genetic Testing for PTEN Hamartoma Tumor Syndrome
- Moderate Penetrance Variants Associated With Breast Cancer in Individuals at High Breast Cancer Risk
- Use of Common Genetic Variants (Single Nucleotide Variants) to Predict Risk of Nonfamilial Breast Cancer

### Benefit Application

Benefit determinations should be based in all cases on the applicable contract language. To the extent there are any conflicts between these guidelines and the contract language, the contract language will control. Please refer to the member's contract benefits in effect at the time of service to determine coverage or non-coverage of these services as it applies to an individual member.
Some state or federal mandates (e.g., Federal Employee Program [FEP]) prohibits plans from denying Food and Drug Administration (FDA)-approved technologies as investigational. In these instances, plans may have to consider the coverage eligibility of FDA-approved technologies on the basis of medical necessity alone.

### Regulatory Status

Clinical laboratories may develop and validate tests in-house and market them as a laboratory service; laboratory-developed tests must meet the general regulatory standards of the Clinical Laboratory Improvement Amendments. Laboratories that offer laboratory-developed tests must be licensed by the Clinical Laboratory Improvement Amendments for high-complexity testing. To date, the U.S. Food and Drug Administration has chosen not to require any regulatory review of these tests.

### Rationale

#### Background

**Genetic Testing for Cancer Susceptibility**

Genetic testing for cancer susceptibility may be approached by a focused method that involves testing for well-characterized variants based on a clinical suspicion of which gene(s) may be the cause of the heritable or familial cancer. Panel testing involves evaluating for multiple variants in multiple genes at one time.

Multiple commercial companies and medical center laboratories offer genetic testing panels that use next-generation sequencing (NGS) methods for hereditary cancers. NGS is one of several methods that use massively parallel platforms to allow the sequencing of large stretches of DNA. Panel testing is potentially associated with greater efficiencies in the evaluation of genetic diseases; however, it may provide information on genetic variants of uncertain clinical significance or findings that would not lead to changes in patient management. Currently available panels do not include all genes associated with hereditary cancer syndromes. Also, these panels may not test for variants (i.e., single nucleotide variants), which may be associated with a low, but increased cancer risk.

#### Genes Included in NGS Panels

The following summarizes the function and disease association of major genes included in NGS panels. This summary is not comprehensive.

**BRCA1 and BRCA2 Variants**

BRCA1 and BRCA2 germline variants are associated with hereditary breast and ovarian cancer syndrome, which is associated most strongly with increased susceptibility to breast cancer at an early age, bilateral breast cancer, male breast cancer, ovarian cancer, cancer of the fallopian tube, and primary peritoneal cancer. BRCA1 and BCRA2 variants are also associated with increased risk of other cancers, including prostate cancer, pancreatic cancer, gastrointestinal cancers, melanoma, and laryngeal cancer.

**APC Variants**

APC germline variants are associated with familial adenomatous polyposis (FAP) and attenuated FAP. FAP is an autosomal dominant colon cancer predisposition syndrome characterized by hundreds to thousands of colorectal adenomatous polyps and accounts for about 1% of all colorectal cancers (CRCs).

**ATM Variants**

ATM is associated with the autosomal recessive condition ataxia-telangiectasia. This condition is characterized by progressive cerebellar ataxia with onset between the ages of 1 and 4 years,
telangiectasias of the conjunctivae, oculomotor apraxia, immune defects, and cancer predisposition, particularly leukemia and lymphoma.

**BARD1, BRIP1, MRE11A, NBN, RAD50, and RAD51C Variants**

BARD1, BRIP1, MRE11A, NBN, RAD50, and RAD51C are genes in the Fanconi anemia/BRCA pathway. Variants in these genes are estimated to confer up to a 4-fold increase in the risk for breast cancer. This pathway is also associated with a higher risk of ovarian cancer and, less often, pancreatic cancer.

**BMPR1A and SMAD4 Variants**

BMPR1A and SMAD4 are genes mutated in juvenile polyposis syndrome and account for 45% to 60% of cases of juvenile polyposis syndrome. Juvenile polyposis syndrome is an autosomal dominant disorder that predisposes to the development of polyps in the gastrointestinal tract. Malignant transformation can occur, and the risk of gastrointestinal cancer has been estimated from 9% to 50%.

**CHEK2 Variants**

CHEK2 gene variants confer an increased risk of developing several different types of cancer, including breast, prostate, colon, thyroid, and kidney. CHEK2 regulates the function of the BRCA1 protein in DNA repair and has been associated with familial breast cancers.

**CDH1 Variants**

CDH1 germline variants are associated with lobular breast cancer in women and with hereditary diffuse gastric cancer (DGC). The estimated cumulative risk of gastric cancer for CDH1 variant carriers by age 80 years is 70% for men and 56% for women. CDH1 variants are associated with a lifetime risk of 39% to 52% of lobular breast cancer.

**EPCAM, MLH1, MSH2, MSH6, and PMS2 Variants**

EPCAM, MLH1, MSH2, MSH6, and PMS2 are mismatch repair genes associated with Lynch syndrome (hereditary nonpolyposis CRC). Lynch syndrome is estimated to cause 2% to 5% of all colon cancers. Lynch syndrome is associated with a significantly increased risk of several types of cancer—colon cancer (60%-80% lifetime risk), uterine/endometrial cancer (20%-60% lifetime risk), gastric cancer (11%-19% lifetime risk), and ovarian cancer (4%-13% lifetime risk). The risks of other types of cancer, including small intestine, hepatobiliary tract, upper urinary tract, and brain, are also elevated.

**MUTYH Variants**

MUTYH germline variants are associated with an autosomal recessive form of hereditary polyposis. It has been reported that 33% and 57% of patients with clinical FAP and attenuated FAP, respectively, who are negative for variants in the APC gene, have MUTYH variants.

**PALB2 Variants**

PALB2 germline variants are associated with an increased risk of pancreatic and breast cancer. Familial pancreatic and/or breast cancer due to PALB2 variants is inherited in an autosomal dominant pattern.

**PTEN Variants**

PTEN variants are associated with PTEN hamartoma tumor syndrome (PHTS), which includes Cowden syndrome (CS), Bannayan-Riley-Ruvalcaba syndrome, and Proteus syndrome. CS is characterized by a high risk of developing tumors of the thyroid, breast, and endometrium. Affected persons have a lifetime risk of up to 50% for breast cancer, 10% for thyroid cancer, and 5% to 10% for endometrial cancer.
**STK11 Variants**
STK11 germline variants are associated with Peutz-Jeghers syndrome, an autosomal dominant disorder, with a 57% to 81% risk of developing cancer by age 70, of which gastrointestinal and breast cancers are the most common.

**TP53 Variants**
TP53 are associated with Li-Fraumeni syndrome. People with TP53 variants have a 50% risk of developing any of the associated cancers by age 30 and a lifetime risk up to 90%, including sarcomas, breast cancer, brain tumors, and adrenal gland cancers.

**NF1 Variants**
Neurofibromin 1 (NF1) encodes a negative regulator in the ras signal transduction pathway. Variants in the NF1 gene have been associated with neurofibromatosis type 1, juvenile myelomonocytic leukemia, and Watson syndrome.

**RAD51D Variants**
RAD51D germline variants are associated with familial breast and ovarian cancers.

**CDK4 Variants**
Cyclin-dependent kinase-4 (CDK4) is a protein-serine kinase involved in cell cycle regulation. Variants in this gene are associated with a variety of cancers, particularly cutaneous melanoma.

**CDKN2A Variants**
Cyclin-dependent kinase inhibitor 2A (CDKN2A) encodes proteins that act as multiple tumor suppressors through their involvement in 2 cell cycle regulatory pathways: the p53 pathway and the RB1 pathway. Variants or deletions in CDKN2A are frequently found in multiple types of tumor cells. Germline variants in CDKN2A have been associated with risk of melanoma, along with pancreatic and central nervous system cancers.

**RET Variants**
RET encodes a receptor tyrosine kinase; variants in this gene are associated with multiple endocrine neoplasia syndromes (types IIA and IIB) and medullary thyroid carcinoma.

**SDHA, SDHB, SDHC, SDHD, and SDHAF2 Variants**
SDHA, SDHB, SDHC, SDHD, and SDHAF2 gene products are involved in the assembly and function of 1 component of the mitochondrial respiratory chain. Germline variants in these genes are associated with the development of paragangliomas, pheochromocytomas, gastrointestinal stromal tumors, and a PTEN-negative Cowden-like syndrome.

**TMEM127 Variants**
Transmembrane protein 127 (TMEM127) germline variants are associated with risk of pheochromocytomas.

**VHL Variants**
VHL germline variants are associated with Hippel-Lindau syndrome, an autosomal dominant familial cancer syndrome. This syndrome is associated with various malignant and benign tumors, including central nervous system tumors, renal cancers, pheochromocytomas, and pancreatic neuroendocrine tumors.

**FH Variants**
Fumarate hydratase (FH) variants are associated with renal cell and uterine cancers.

**FLCN Variants**
Folliculin (FLCN) acts as a tumor suppressor gene; variants in this gene are associated with the autosomal dominant Birt-Hogg-Dube syndrome, which is characterized by hair follicle hamartomas, kidney tumors, and CRC.
MET Variants
MET is a proto-oncogene that acts as the hepatocyte growth factor receptor. MET variants are associated with hepatocellular carcinoma and papillary renal cell carcinoma.

MITF Variants
Microphthalmia-associated transcription factor (MITF) is a transcription factor involved in melanocyte differentiation. MITF variants lead to several auditory-pigmentary syndromes, including Waardenburg syndrome type 2 and Tietze syndrome. MITF variants are also associated with melanoma and renal cell carcinoma.

TSC1 Variants
Tuberous sclerosis 1 (TSC1) and tuberous sclerosis 2 (TSC2) encode the proteins hamartin and tuberin, which are involved in cell growth, differentiation, and proliferation. Variants in these genes are associated with the development of tuberous sclerosis complex, an autosomal dominant syndrome characterized by skin abnormalities, developmental delay, seizures, and multiple types of cancers, including central nervous system tumors, renal tumors (including angiomyolipomas, renal cell carcinomas), and cardiac rhabdomyomas.

XRCC2 Variants
XRCC2 encodes proteins thought to be related to the RAD51 protein product that is involved in DNA double-stranded breaks. Variants may be associated with Fanconi anemia and breast cancer.

FANCC Variants
Fanconi anemia complementation group C (FANCC) is one of several DNA repair genes that mutate in Fanconi anemia, which is characterized by bone marrow failure and a high predisposition to multiple types of cancer.

AXIN2 Variants
AXIN2 variants are associated with FAP syndrome, although the phenotypes associated with AXIN2 variants do not appear to be well-characterized.

Hereditary Cancer and Cancer Syndromes
Genetic testing for breast and ovarian cancer syndromes, single nucleotide variants related to breast cancer, and hereditary breast cancer are evaluated in Blue Shield of California Medical Policies: Genetic Testing for Hereditary Breast/Ovarian Cancer Syndrome (BRCA1 or BRCA2), Use of Common Genetic Variants (Single Nucleotide Variants) to Predict Risk of Nonfamilial Breast Cancer, Moderate Penetrance Variants Associated With Breast Cancer in Individuals at High Breast Cancer Risk, respectively.

Genetic testing for Li-Fraumeni syndrome is evaluated in Blue Shield of California Medical Policy: Genetic Testing for Li-Fraumeni Syndrome.

CS is a part of PHTS and is the only PHTS disorder associated with a documented predisposition to malignancies. Genetic testing for CS is evaluated in Blue Shield of California Medical Policy: Genetic Testing for PTEN Hamartoma Tumor Syndrome.

Hereditary Diffuse Gastric Cancer
Hereditary DGC is an autosomal dominant trait. Up to 50% of familial cases may be caused by variants in the CDH1 gene. In kindred families with CDH1-positive hereditary DGC, the risk of developing DGC is as high as 80% by 80 years of age. Other candidate genes include CTNNA1, BRCA2, STK11, SDHB, PRSS1, ATM, MSR1, and PALB2. Guidelines from the International Gastric Cancer Linkage Consortium have proposed genetic testing in families with 2 or more patients with gastric cancer at any age, in individuals with DGC before the age of 40, or in families with diagnoses of both DGC and invasive lobular cancer. Because of the high lifetime risk, prophylactic total gastrectomy between the ages of 20 and 30 is usually advised.
Hereditary Colon Cancer Syndromes

Genetic testing for hereditary colon cancer syndromes are addressed in a related policy (see Blue Shield of California Medical Policy: Genetic Testing for Lynch Syndrome and Other Inherited Colon Cancer Syndromes). Hereditary colon cancer syndromes are thought to account for approximately 10% of all CRCs. Another 20% have a familial predilection for CRC without a clear hereditary syndrome identified. The hereditary CRC syndromes can be divided into the polyposis and nonpolyposis syndromes. Although there may be polyps in the nonpolyposis syndromes, they are usually less numerous; the presence of 10 colon polyps is used as a rough threshold when considering genetic testing for a polyposis syndrome. The polyposis syndromes can be further subdivided by polyp histology, which includes the adenomatous (FAP, attenuated FAP, MUTYH-associated) and hamartomatous (juvenile polyposis syndrome, Peutz-Jeghers syndrome, PHTS) polyposis syndromes. The nonpolyposis syndromes include Lynch syndrome.

Literature Review

Evidence reviews assess whether a medical test is clinically useful. A useful test provides information to make a clinical management decision that improves the net health outcome. That is, the balance of benefits and harms is better when the test is used to manage the condition than when another test or no test is used to manage the condition.

The first step in assessing a medical test is to formulate the clinical context and purpose of the test. The test must be technically reliable, clinically valid, and clinically useful for that purpose. Evidence reviews assess the evidence on whether a test is clinically valid and clinically useful. Technical reliability is outside the scope of these reviews, and credible information on technical reliability is available from other sources.

Cancer Susceptibility Panels

Cancer susceptibility panels may be either diagnostic or prognostic.

Clinical Context and Test Purpose

The purpose of diagnostic testing symptomatic patients for genetic or heritable pathogenic variants is to establish a molecular diagnosis defined by the presence of a known pathologic variant(s). For genetic testing, a symptomatic individual is defined as one with a clinical phenotype that correlates with a known pathologic variant but who has not yet developed a malignancy. The criteria under which testing for genetic cancer susceptibility may be considered clinically useful are as follows:

- An association of the marker with the disorder has been established;
- Symptoms of the disease are present;
- A definitive diagnosis cannot be made based on history, physical examination, pedigree analysis, and/or standard diagnostic studies or tests; and
- The clinical utility of diagnosis has been established (e.g., by demonstrating that a definitive diagnosis will lead to changes in the clinical management of the condition, changes in surveillance, or changes in reproductive decision making, and these changes will lead to improved health outcomes).

The purpose of prognostic testing for cancer susceptibility is to predict whether a cancer is likely to occur in a family member of an affected person. The criteria under which prognostic testing may be considered clinically useful are as follows:

- An association of the marker with the natural history of the disease has been established; and
- The clinical utility of identifying the variant has been established (e.g., by demonstrating that testing will lead to changes in the clinical management of the condition or changes in surveillance).

The question addressed in this evidence review is: Does testing for genetic cancer susceptibility improve the net health outcome?
The following PICOTS were used to select literature to inform this review.

**Patients**
The relevant population of interest are patients being evaluated for clinical signs or symptoms that may be associated with a risk for the presence of a heritable cancer variant (diagnostic testing) or have a family member(s) diagnosed with heritable cancer(s) (prognostic testing).

**Intervention**
The test being considered is a cancer susceptibility panel.

**Comparator**
The following test is currently being used to make decisions about managing cancer susceptibility: individual gene variant testing.

**Outcomes**
The outcomes of interest are sensitivity and specificity, positive and negative predictive value, and reductions in morbidity and mortality.

**Timing**
Follow-up varies by whether testing is diagnostic or prognostic.

**Setting**
These tests are offered commercially through various manufacturers and institutions.

**Technically Reliable**
Assessment of technical reliability focuses on specific tests and operators and requires a review of unpublished and often proprietary information. Review of specific tests, operators, and unpublished data are outside the scope of this evidence review and alternative sources exist. This evidence review focuses on the clinical validity and clinical utility.

**Clinically Valid**
A test must detect the presence or absence of a condition, the risk of developing a condition in the future, or treatment response (beneficial or adverse).

The published literature provides no guidance on the assessment of the clinical validity of panel testing for cancer susceptibility with next-generation sequencing (NGS), and the usual approach to establishing the clinical validity of genetic testing is difficult to apply to panel testing.

Although it may be possible to evaluate the clinical validity of the sequencing of individual genes found on these panels, the clinical validity of NGS for cancer susceptibility panels, which include variants associated with unknown or variable cancer risk, are of uncertain clinical validity.

For genetic susceptibility to cancer, clinical validity can be considered at the following levels:
1. Does a positive test identify a person as having an increased risk of developing cancer?
2. If so, how high is the risk of cancer associated with a positive test?

**Hereditary Cancer Panels**
The likelihood that someone with a positive test result will develop cancer is affected not only by the presence of the gene variant but also by other modifying factors that can affect the penetrance of the variant (e.g., environmental exposures, personal behaviors) or by the presence or absence of variants in other genes.

Susswein et al (2016) reviewed the genetic test results and clinical data from a consecutive series of 10030 patients referred for evaluation by a hereditary cancer panel between August 2013 and October 2014. Personal and family histories of cancer were obtained, and patients
genes included in the panels were grouped into 3 risk categories based on penetrance data available in 2012, as follows:
- High risk: APC, BMPR1A, BRCA1, BRCA2, CDH1, CDKN2A, EPCAM, MLH1, MSH2, MSH6, MUT YH, PMS2, PTEN, SMAD4, STK11, TP53, and VHL
- moderate risk: ATM, CHEK2, and PALB2
- increased but less well defined risk: AXIN2, BARD1, BRIP1, CDK4, FANCC, NBN, RAD51C, RAD51D, and XRCC2.

Over half of the individuals referred for testing were women with breast cancer (n=5209), of whom 3315 (63.6%) had not had previous BRCA1 and BRCA2 testing. Unaffected individuals comprised 25.2% of the study population. Overall, 9.0% (901/10030) of the patients were found to carry at least 1 pathogenic or likely pathogenic variant, totaling 937 variants. Approximately half of the positive results were in well-established genes (including BRCA1 and BRCA2, Lynch syndrome, and other high-risk genes) and approximately half in genes with moderate or unknown risk. Likely pathogenic variants comprised 10.6% (99/937) of all positive results, with CHEK2 accounting for the majority of all likely pathogenic variants (68.7% [68/99]).

Individuals with colon/stomach cancer had the highest yield of positive results (14.8% [113/764]), the majority of which were in well-established colon cancer genes: MLH1, MSH2, MSH6, PMS2, EPCAM, MUTYH, APC, PTEN, and STK11. However, 28.2% (35/124) were observed in genes not considered classical for gastrointestinal cancers: BRCA1, BRCA2, CHEK2, ATM, PALB2, BRIP1, and RAD51D. BRCA1 and BRCA2 accounted for 9.7% (12/124) of positive variants identified in individuals diagnosed with colon cancer. The Lynch syndrome/colorectal cancer (CRC) panel (GeneDx), containing MLH1, MSH2, MSH6, PMS2, EPCAM, APC, and MUTYH, had the highest yield (13.7% overall; 17.6% among affected individuals). The panel’s high yield was likely a result of the well-established association of all genes on this panel with CRC and the specific clinical history or tumor characteristics (microsatellite instability and/or immunohistochemistry) that prompted providers to order this focused panel.

The breast cancer high-risk panel containing BRCA1, BRCA2, CDH1, PTEN, STK11, and TP53 had the lowest yield (3.8% overall, 4.2% among individuals with breast cancer). The highest VUS frequency was observed with the largest panel (29 genes), and the lowest VUS rate was observed with the high-risk breast cancer panel with 6 genes. For patients with breast cancer, 9.7% (320/3315) of women without prior BRCA1 and BRCA2 testing were found to carry a pathogenic or likely pathogenic variant, of which BRCA1 and BRCA2 accounted for 39.1%. Other high-risk genes (including TP53, PTEN, and CDH1) accounted for 5.8% (19/330), and 5.2% (17/330) of the patients carried the Lynch syndrome genes. Moderate and less well-defined risk genes accounted for 50.0% (165/330) of all positive results among women with breast cancer. Of women with ovarian cancer without previous BRCA1 and BRCA2 testing, 13.4% (89/663) had variants, of which BRCA1 and BRCA2 accounted for 50.5% Lynch syndrome genes for 14.3% and moderate or less well-defined risk genes for 33.0%. Of the 453 women with endometrial cancer, the yield for identifying a variant was 11.9% (n=54): 7.3% (n=33) of these were within a Lynch gene, most commonly MSH6; CHEK2 was positive in 7%, with an overall frequency of 1.5% and 6 positive results were identified in BRCA1 and BRCA2, 10.9% (6/55) of all positive variants identified.
Among 190 pancreatic cancer patients, the yield for identifying a variant was 10.5% (n=20), most commonly identified in ATM (40.0% [8/20]), BRCA2 (25.0% [5/20]), and PALB2 (15.0% [3/20]). Of 901 patients with positive results, 28 (3.1%) had more than 1 positive finding, reflecting 0.3% (28/10030) of the total testing population; 5 had positive results in 2 highly penetrant genes; 12 had 1 positive result in a high-risk gene, and 1 in a gene with moderate or unknown risk; and 11 had 2 positive findings in genes with moderate or unknown risk.

Six (33%) of the 18 patients with positive findings in TP53 did not meet classic Li-Fraumeni syndrome, Li-Fraumeni-like syndrome, 2009 Chompret, or National Comprehensive Cancer Network guideline criteria for TP53 testing, resulting in a frequency of 0.06% (6/9605) unanticipated positive results. Four patients had a positive CDH1 result, two of whom did not meet the International Gastric Cancer Linkage Consortium testing criteria, resulting in a frequency of 0.02% (2/8708) positive CDH1 results. In summary, among patients with specific cancers, yields were 9.7%, 13.4%, and 14.8% in patients with breast, ovarian, and colon/stomach cancers, respectively. Approximately 5.8% of positive results among women with breast cancer were in highly penetrant genes other than BRCA1 and BRCA2. The yield in Lynch syndrome genes among breast cancer patients was 0.5% (17/3315), higher than a published upper estimate of the prevalence of Lynch among the general population (0.2%). More than a quarter of patients with colon cancer tested positive for genes not considered to be classic CRC genes. Over 11% of the positive findings among women with endometrial cancer were in BRCA1 and BRCA2. A small number of patients whose personal and family histories were not suggestive of Li-Fraumeni syndrome were positive for pathogenic variants in the TP53 gene.

LaDuca et al (2014) reported on the clinical and molecular characteristics of 2079 patients who underwent panel testing with BreastNext, OvaNext, ColoNext, or CancerNext (Ambry Genetics).5 Most (94%) patients had a personal history of cancer or adenomatous polyps, and in 5% of cases, the proband was reported to be clinically unaffected. A total of 2079 cases were included: 874 BreastNext, 222 OvaNext, 557 ColoNext, and 425 CancerNext. The positive and inconclusive rates for the panels were, respectively, 7.4% and 20% for BreastNext, 7.2% and 26% for OvaNext, 9.2% and 15% for ColoNext, and 9.6% and 24% for CancerNext.

**Hereditary Breast and Ovarian Cancer**

O’Leary et al (2017) reported on 1085 cases with non-BRCA1 or BRCA2 breast cancer referred to a commercial laboratory that were found to have a pathogenic or likely pathogenic variant.6 The cases were divided into three groups based on the panel requested by the ordering physician: genes primarily associated with breast cancer (group A), genes associated with breast, gynecologic, and gastrointestinal cancer types (group B), and large comprehensive panels (group C). The proportion of positive finding in genes with breast management guidelines was inversely related to the size of the panel: 97.5% in group A, 63.6% in group B, and 50% in group C. Conversely, more positive findings and unexpected findings (there was no family history) were identified in actionable non breast cancer genes as the size of the panel increased. VUS rates also increased as the size of the panel increased, with 12.7% VUS in group A, 31.6% in group B, and 49.6% in group C.

Couch et al (2017) evaluated 21 genetic predisposition genes for breast cancer in a sample of 38326 white women with breast cancer who received any one of a variety of genetic test panels (Ambry Genetics).7 The frequency of pathogenic variants was estimated at 10.2%. After the exclusion of BRCA1, BRCA2, and syndromic breast cancer genes (CDH1, PTEN, TP53), five additional genes with variants classified as pathogenic by ClinVar were associated with high or moderately increased risk of breast cancer (see Table 3). Notably, of the various panels included in this study, only the BRCA plus panel is limited to the set of genes (ATM, BRCA1, BRCA2, CDH1, CHEK2, PALB2, PTEN) that were associated with breast cancer in women of European descent.
Table 1. Moderate-to-High Risk Non-BRCA1 and BRCA2, Nonsyndromic Genes Associated With Breast Cancer

<table>
<thead>
<tr>
<th>Gene</th>
<th>Odds Ratio</th>
<th>95% Confidence Interval</th>
<th>Risk Category</th>
</tr>
</thead>
<tbody>
<tr>
<td>ATM</td>
<td>2.78</td>
<td>2.22 to 3.62</td>
<td>Moderate</td>
</tr>
<tr>
<td>BARD1</td>
<td>2.16</td>
<td>1.31 to 3.63</td>
<td>Moderate</td>
</tr>
<tr>
<td>CHEK2</td>
<td>1.48</td>
<td>1.31 to 1.67</td>
<td>Moderate</td>
</tr>
<tr>
<td>PALB2</td>
<td>7.46</td>
<td>5.12 to 11.19</td>
<td>High</td>
</tr>
<tr>
<td>RAD51D</td>
<td>3.07</td>
<td>1.21 to 7.88</td>
<td>Moderate</td>
</tr>
</tbody>
</table>

Other studies have assessed the prevalence of pathogenic variants among patients with breast cancer who were referred for genetic testing, using a panel of 25 genes associated with inherited cancer predisposition (Myriad Genetics). A study by Buys et al (2017) included over 35000 women with breast cancer who were assessed with the Myriad 25-gene panel. Pathogenic variants were identified in 9.3% of the women tested. Nearly half of those variants were in the BRCA1 or BRCA2 genes. The remaining variants were found in other breast cancer genes, Lynch syndrome genes, and other panel genes. The VUS rate was 36.7%

A similar study by Langer et al (2016) evaluated the frequency of pathogenic variants identified with the 25-gene panel (Myriad Genetics) in 3088 patients with a personal history of ovarian cancer who were referred for testing. Pathogenic or likely pathogenic variants were identified in 419 (13.6%) patients, of whom 7 patients had variants in 2 different genes. Nearly all patients (99.2%) met National Comprehensive Cancer Network guidelines for hereditary breast and ovarian cancer testing (78.4%), Lynch syndrome testing (0.3%), or both (20.5%). Of the 419 patients with pathogenic or likely pathogenic variants, 277 (65%) were identified in BRCA1 or BRCA2: 33 (7.8%) in Lynch syndrome-associated genes (PMS2, MSH6, MLH1, MSH2), and 26.8% in genes with a low-to-moderate increase in cancer risk (ATM, BRIP1, CHEK2, RAD51C, PALB2, NBN), or one of 6 other genes (<1% each). One or more VUS were reported in 1141 (36.9%) of patients.

Tung et al (2015) included 2 cohorts: 1781 patients referred for commercial testing for BRCA1 and BRCA2 variants and whose samples were consecutively submitted to Myriad between November 2012 and April 2013 (cohort 1), and 377 DNA samples from patients who were referred to Beth Israel Deaconess Medical Center for genetic testing between 1998 and 2013 and had previously tested negative for BRCA1 and BRCA2 (cohort 2). Variants were identified in 16 genes, with the most frequent being BRCA1, BRCA2, CHEK2, ATM, and PALB2.

Colorectal Cancer
Hansen et al (2017) published a retrospective analysis using multigene panel testing to identify genetic causes for increased CRC risk. A custom gene panel targeting 112 genes, including both well-known and candidate CRC susceptibility genes, was designed, and variants were validated by Sanger sequencing. DNA samples from 274 familial CRC patients who fulfilled the Amsterdam I/II and/or the Revised Bethesda guidelines were included. All had previously been screened for variants in 1 or more of the MMR genes (MLH1, MSH2, MSH6, PMS2, EPCAM, BMPR1, SMAD4, STK11, APC, MUTYH, CHEK2, TP53, PTEN, and CDH1 genes. Alterations were classified as follows: (1) pathogenic variant; (2) variant, likely pathogenic; (3) variant, unknown significance; (4) variant, likely benign; and (5) benign. Data were analyzed for 586 patients whose ColoNext testing results and associated clinical data were maintained in a database by Ambry Genetics. Sixty-one (10.4%) patients had genetic alterations consistent with pathogenic variants or likely pathogenic variants; after 8 patients with only CHEK2 or 1 MUTYH variant were removed, 42 (7.2%) patients were considered to have
actionable variants. One hundred eighteen (20.1%) patients had at least 1 VUS, including 14 patients who had at least 1 VUS in addition to a pathologic variant. Of the 42 patients with a pathologic variant, most (30 [71%] patients) met National Comprehensive Cancer Network guidelines for syndrome-based testing, screening, or diagnosis, based on the available clinical and family history. The authors noted “The reality remains that syndrome based testing would have been sufficient to identify the majority of patients with deleterious variants. Consequently, the optimal and most cost-effective use of panel-based testing as a first-tier test vsa second-tier test (i.e. after syndrome-based testing is negative), remains to be determined.”

Section Summary: Clinically Valid
Clinical validity studies have studied mixed populations; high-risk individuals due to clinical presentation or family history as well as cancer-affected persons with or without prior variant testing. Most studies have been retrospective. These studies have reported on the frequency with which well-known cancer susceptibility variants are identified using large panels and variably have reported the VUS rate. VUS rates increased in proportion with panel size, reaching nearly 50% for large gene panels.

Clinically Useful
A test is clinically useful if the use of the results informs management decisions that improve the net health outcome of care. The net health outcome can be improved if patients receive correct therapy, or more effective therapy, or avoid unnecessary therapy, or avoid unnecessary testing.

Direct Evidence
Direct evidence of clinical utility is provided by studies that have compared health outcomes for patients managed with and without the test. Because these are intervention studies, the preferred evidence would be from randomized controlled trials.

Chain of Evidence
Indirect evidence on clinical utility rests on clinical validity. If the evidence is insufficient to demonstrate test performance, no inferences can be made about clinical utility.

The following criteria can be used to evaluate the clinical utility of cancer susceptibility panel testing:
- Does panel testing offer substantial advantages in efficiency compared with sequential analysis of individual genes?
- Is decision making based on potential results of panel testing well-defined?
  o Do positive results on panel testing result in changes in cancer susceptibility that are clinically important?
  o Does this change in cancer susceptibility lead to changes in management that result in health outcome benefits for the patient being tested?
- Is the impact of ancillary information provided by panel testing well-defined?
  o What is the probability that ancillary information leads to further testing or management changes that may have either a positive or a negative impact on the patient being tested?

Identifying a person with a genetic variant that confers a high-risk of developing cancer could lead to changes in clinical management and improve health outcomes. There are well-defined clinical guidelines on the management of patients who are identified as having high-risk hereditary cancer syndrome. Changes in clinical management could include modifications in cancer surveillance, specific risk-reducing measures (e.g., prophylactic surgery), and treatment guidance (e.g., avoidance of certain exposures). Also, other at-risk family members could be identified.

On the other hand, identifying variants that have intermediate or low penetrance is of limited clinical utility. Clinical management guidelines for patients found to have one of these variants
are not well-defined. Also, there is a potential for harm, in that the diagnosis of an intermediate-
or low-risk variant may lead to undue psychological stress and unnecessary prophylactic surgical
intervention.

Idos et al (2018) conducted a prospective study that enrolled 2000 patients who had been
referred for genetic testing at 1 of 3 academic medical centers (see Table 2). Patients
underwent differential diagnosis by a genetic clinician prior to cancer panel testing for 25 or 28
genes associated with breast or ovarian cancer, Lynch syndrome, and genes associated with
gastric, colon, or pancreatic cancer. Results of the study are shown in Table 3. Twelve percent of
the patients were found to have a pathogenic variant, of which 66% was anticipated by the
genetic clinician and 34% which were not anticipated. Most of the unanticipated results were in
moderate to low penetrance genes. Thirty-four percent of the patients had a VUS and 53% of
patients had benign results. Prophylactic surgery was performed more frequently in patients with
a pathogenic variant (16%) compared to patients with a benign (2.4%) or unknown (2.3%)
variant. Limitations in relevance and design and conduct are shown in Tables 4 and 5.

Information on the actions associated with low to moderate penetrance genes were not
reported. One concern with large panels is the increase in VUS. Having a VUS did not increase
distress or uncertainty or diminish a positive experience of the testing in this study, and there was
no increase in prophylactic surgery in patients with a VUS. However, all patients had received
genetic counseling at an academic medical center regarding the outcomes of testing and this
study may not be representative of community practice. In addition, a threshold for testing of
2.5% on a risk prediction model is a lower threshold than what is typically recommended.
Patients with a positive result were more likely to encourage relatives to undergo testing. Longer-
term follow-up for clinical outcomes is ongoing.

Table 2. Study Characteristics

<table>
<thead>
<tr>
<th>Study</th>
<th>Study Population</th>
<th>Design</th>
<th>Comparator</th>
<th>Outcomes</th>
<th>Blinding of Assessors</th>
<th>Follow-up</th>
</tr>
</thead>
<tbody>
<tr>
<td>Idos et al (2018)</td>
<td>2,000 patients who underwent a multi-gene cancer panel test</td>
<td>Prospective</td>
<td>Differential diagnosis by a genetic clinician</td>
<td>Post-test survey of decisions and attitudes</td>
<td>No</td>
<td>1,573 surveys were returned at a median of 13 mo after the genetic test</td>
</tr>
</tbody>
</table>

* Patients met genetic testing guidelines or had at least a 2.5% risk of cancer on a risk prediction model. Seventy-three percent had a personal history of cancer. Reasons for genetics referral included cancer diagnosis < 50 years of age, ≥ 2 close relatives with cancer, ≥ 1 family member with cancer at < 50 years of age, or history of multiple cancers.

Table 3. Study Results

<table>
<thead>
<tr>
<th>Study</th>
<th>Initial N</th>
<th>Final N</th>
<th>Clinically Anticipated n (%)</th>
<th>Test Results not Clinically Anticipated n (%)</th>
<th>Outcome n (%)</th>
<th>P-value Pathogenic vs VUS</th>
</tr>
</thead>
<tbody>
<tr>
<td>Overall</td>
<td>62</td>
<td>30 (16.0)</td>
<td>12 (2.3)</td>
<td>20 (2.4)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Prophylactic surgery</td>
<td>1,248</td>
<td>6.1 (6.04)</td>
<td>2.1 (4.2)</td>
<td>1.7 (3.5)</td>
<td>&lt;0.001</td>
<td></td>
</tr>
<tr>
<td>Distress score (0 to 30) - mean (SD)</td>
<td>1,223</td>
<td>11.4 (8.8)</td>
<td>7.4 (7.8)</td>
<td>6.3 (7.1)</td>
<td>&lt;0.001</td>
<td></td>
</tr>
<tr>
<td>Uncertainty (0 to 45) - mean (SD)</td>
<td>1,213</td>
<td>9.7 (5.1)</td>
<td>11.8 (6.3)</td>
<td>12.1 (6.5)</td>
<td>&lt;0.001</td>
<td></td>
</tr>
</tbody>
</table>
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Genetic Cancer Susceptibility Panels Using Next-Generation Sequencing

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(0 to 20) mean (SD)

SD: standard deviation; VUS: variant of uncertain significance.

a 31% had a variant in BRCA1/BRCA2, 16% had a variant associated with Lynch syndrome, 18% had a pathogenic MUTYH variant, and 8% had pathogenic variants in APC. Other genes included TP53, CHEK2, ATM, PALB2, BRIP1, RAD51C, BARD1, NBN, CDH1, and CDKN2A.

Table 4. Relevance Limitations

<table>
<thead>
<tr>
<th>Study</th>
<th>Population</th>
<th>Intervention</th>
<th>Comparator</th>
<th>Outcomes</th>
<th>Follow-Up</th>
</tr>
</thead>
<tbody>
<tr>
<td>Idos et al (2018)</td>
<td>4. The population included patients down to 2.5% of risk on a risk prediction model.</td>
<td>1. The outcomes were patient-reported experience.</td>
<td>1. Follow-up is continuing for clinical outcomes.</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

The study limitations stated in this table are those notable in the current review; this is not a comprehensive limitations assessment.

a Population key: 1. Intended use population unclear; 2. Clinical context is unclear; 3. Study population is unclear; 4. Study population not representative of intended use.

b Intervention key: 1. Not clearly defined; 2. Version used unclear; 3. Delivery not similar intensity as comparator; 4. Not the intervention of interest.

c Comparator key: 1. Not clearly defined; 2. Not standard or optimal; 3. Delivery not similar intensity as intervention; 4. Not delivered effectively.

d Outcomes key: 1. Key health outcomes not addressed; 2. Physiologic measures, not validated surrogates; 3. No CONSORT reporting of harms; 4. Not establish and validated measurements; 5. Clinical significant difference not prespecified; 6. Clinical significant difference not supported.

e Follow-Up key: 1. Not sufficient duration for benefit; 2. Not sufficient duration for harms.

Table 5. Study Design and Conduct Limitations

<table>
<thead>
<tr>
<th>Study</th>
<th>Selection</th>
<th>Blinding</th>
<th>Delivery of Test</th>
<th>Selective Reporting</th>
<th>Data Completeness</th>
<th>Statistical</th>
</tr>
</thead>
<tbody>
<tr>
<td>Idos et al (2018)</td>
<td>1. Blinding not described</td>
<td>1. Surveys were completed by 69% of patients at 3 mo and 57% at 12 mo.</td>
<td>1. Surveys were completed by 69% of patients at 3 mo and 57% at 12 mo.</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

The study limitations stated in this table are those notable in the current review; this is not a comprehensive limitations assessment.

a Selection key: 1. Selection not described; 2. Selection not random or consecutive (ie, convenience).

b Blinding key: 1. Not blinded to results of reference or other comparator tests.

c Test Delivery key: 1. Timing of delivery of index or reference test not described; 2. Timing of index and comparator tests not same; 3. Procedure for interpreting tests not described; 4. Expertise of evaluators not described.


e Data Completeness key: 1. Inadequate description of indeterminate and missing samples; 2. High number of samples excluded; 3. High loss to follow-up or missing data.

f Statistical key: 1. Confidence intervals and/or p values not reported; 2. Comparison with other tests not reported.

Lumish et al (2017) evaluated the impact of hereditary breast and ovarian cancer gene panel testing in 232 patients who had undergone gene panel testing after discussion with a genetic counselor. From this sample, 129 patients had a personal history of cancer (11 with a pathogenic or likely pathogenic variant, 14 with a VUS, 104 with normal test results) and 103 had a family history of cancer (14 with a pathogenic or likely pathogenic variant, 20 with a VUS, 69 with normal test results). The greatest impact of test results was for the 14 patients with a family history of breast or ovarian cancer who received a positive (pathogenic or likely pathogenic) test result, leading to greater distress and more frequent screening in 13 patients and prophylactic surgery in 1. Positive test results for the 11 patients with a personal history of cancer influenced their decision about the type of surgery for 4 (36.4%) patients. For the 20 patients with a family history of cancer and a VUS result, distress increased to an intermediate level, and 7 (35%) patients reported that their test result would impact the decision to have additional screening. The authors of this study noted that the VUS rate would increase with the number of genes in a panel and that the choice of a panel would need to optimize the chance of
receiving results with clinical utility while minimizing the chance of results that have disutility and increase anxiety.

Eliade et al (2017) evaluated the clinical actionability of a multigene panel in a cohort of 583 patients with a family history of breast or ovarian cancer. A pathogenic or likely pathogenic BRCA1 or BRCA2 variant was identified in 51 (9%) patients, and a pathogenic or likely pathogenic variant was identified in 10 other genes in the panel for 37 patients. The most frequently mutated genes were CHEK2 (n=12 [2%]), ATM (n=9 [1.5%]), and PALB2 (n=4 [0.6%]). The identification of a pathogenic/likely pathogenic variant in a high-risk gene or in two genes led to a change in surveillance or prophylactic surgery. In patients with a positive finding in a moderate-risk gene, breast magnetic resonance imaging was recommended, while surveillance according to family history was recommended in patients with a negative finding. There was no change in management in the four women with a positive finding in a low-risk gene (BRIP1, BARD1, RAD50). Individuals with a negative finding could not be reassured, given the possibility of a pathogenic or likely pathogenic variant in an as-yet-undiscovered gene.

Rosenthal et al (2017) published an industry-sponsored study evaluating the clinical utility of a 25-gene pan-cancer panel. The analysis included 252223 consecutive individuals, most of whom (92.8%) met testing criteria for hereditary breast and ovarian cancer and/or Lynch syndrome. Pathogenic variants (n=17340) were identified in 17000 (6.7%) patients; the most common pathogenic variants were BRCA1 and BRCA2 (42.2%), other breast cancer genes (32.9%), Lynch syndrome genes (13.2%), and ovarian cancer genes (6.8%). Among individuals who met only hereditary breast and ovarian cancer or Lynch syndrome testing criteria, half of the pathogenic variants found were genes other than BRCA1 and BRCA2 or Lynch syndrome genes, respectively. The study was limited by reliance on providers for personal and family cancer histories and by uncertainty regarding the exact cancer risk spectrum for each gene included on the panel.

Kurian et al (2014) evaluated the information from an NGS panel of 42 cancer-associated genes in women previously referred for clinical BRCA1 and BRCA2 testing after clinical evaluation of hereditary breast and ovarian cancer from 2002 to 2012. The authors aimed to assess concordance of the results of the panel with prior clinical sequencing, the prevalence of potentially clinically actionable results, and the downstream effects on cancer screening and risk reduction. Potentially actionable results were defined as pathogenic variants that cause recognized hereditary cancer syndromes or have a published association with a 2-fold or greater relative risk of breast cancer compared with average-risk women. In total, 198 women participated in the study. Of these, 174 had breast cancer and 57 carried 59 germline BRCA1 and BRCA2 variants. Testing with the panel confirmed 57 of 59 of the pathogenic BRCA1 and BRCA2 variants (n=141), 16 had pathogenic variants in other genes (11.4%). The affected genes were ATM (n=2), BLM (n=1), CDH1 (n=1), CDKN2A (n=1), MLH1 (n=1), MUTYH (n=5), NBN (n=2), PRSS1 (n=1), and SLX4 (n=2). Eleven of these variants had been previously reported in the literature and five were novel. Eighty percent of the women with pathogenic variants in the non-BRCA1 and BRCA2 genes had a personal history of breast cancer. Overall, a total of 428 VUS were identified in 39 genes, among 175 patients. Six women with variants in ATM, BLM, CDH1, NBN, and SLX4 were advised to consider annual breast magnetic resonance imaging because of an estimated doubling of breast cancer risk, and/or and six with variants in CDH1, MLH1, and MUTYH were advised to consider frequent colonoscopy endoscopic gastroduodenoscopy (once every 1-2 years) due to estimated increases in gastrointestinal cancer risk. One patient with an MLH1 variant consistent with Lynch syndrome underwent risk-reducing salpingo-oophorectomy and early colonoscopy, which identified a tubular adenoma that was excised (she had previously undergone hysterectomy for endometrial carcinoma). No clinical outcomes associated with the recommendations were reported.
Chain of Evidence
Indirect evidence on clinical utility rests on clinical validity. If the evidence is insufficient to
demonstrate test performance, no inferences can be made about clinical utility.

Because the clinical validity of cancer susceptibility panel testing for inherited cancer syndromes
has not been established, a chain of evidence cannot be constructed.

Section Summary: Clinically Useful
Data are lacking for the clinical utility of multigene panels for inherited cancer susceptibility
panels. There are management guidelines for syndromes with high penetrance, which have
clinical utility in that they inform clinical decision making and result in the prevention of adverse
health outcomes. Clinical management recommendations for the inherited conditions
associated with low-to-moderate penetrance are not standardized, and the clinical utility of
genetic testing for these variants is uncertain and could potentially lead to harm. Also, high VUS
rates have been reported with the use of these panels.

Summary of Evidence
For individuals who have a personal and/or family history suggesting an inherited cancer
syndrome who receive NGS panel testing, the evidence includes reports describing the
frequency of detecting variants in patients referred for panel testing. The relevant outcomes are
overall survival, disease-specific survival, and test validity. The accuracy of NGS may be
reduced in complex genomic regions, and the interpretation of the significance of the variant
(i.e., pathogenic, benign, or variants of uncertain significance) can differ between laboratories.
Clinical validity studies have reported on the results of the frequency with which variants are
identified. The rates of variants of uncertain significance for gene panels are significant and
increase in proportion with panel size, reaching nearly 50% for large gene panels. Published data
on clinical utility is lacking, and it is unknown whether the use of these panels improves health
outcomes. Variants included in these panels are associated with varying levels of risk of
developing cancer. Only some variants included on panels are associated with a high risk of
developing a well-defined cancer syndrome for which there are established clinical
management guidelines. Many panels include genetic variants considered to be of moderate
or low penetrance, and clinical management recommendations for these genes are not well-
defined. The lack of clinical management pathways for variants of uncertain significance
increases the potential for harm. The evidence is insufficient to determine the effects of the
technology on health outcomes.

Supplemental Information
Practice Guidelines and Position Statements

American Society of Clinical Oncology
The American Society of Clinical Oncology (2015) issued a policy statement on genetic and
genomic testing for cancer susceptibility.18 The update addressed the application of next-
generation sequencing and confirmed that panel testing may also identify variants in genes
associated with moderate or low cancer risks, variants in high-penetrance genes that would not
have been evaluated based on the presenting personal or family history, and as variants of
uncertain significance in a substantial proportion of patient cases. Further, the statement
indicated there is little consensus as to which genes should be included on panels for cancer
susceptibility testing.

National Comprehensive Cancer Network
National Comprehensive Cancer Network guidelines on genetic/familial high-risk assessment for
breast and ovarian cancers (v.3.2019)19 state the following on multigene testing:

- "Patients who have a personal or family history suggestive of a single inherited cancer
  syndrome are most appropriately managed by genetic testing for that specific
  syndrome. When more than one gene can explain an inherited cancer syndrome, then
  multi-gene testing may be more efficient and/or cost-effective."
• There may be a role for multi-gene testing in individuals who have tested negative (indeterminate) for a single syndrome, but whose personal or family history remains suggestive of an inherited susceptibility.
• As commercially available tests differ in the specific genes analyzed (as well as classification of variants and many other factors), choosing the specific laboratory and test panel is important.
• Multi-gene testing can include “intermediate” penetrant (moderate-risk) genes. For many of these genes, there are limited data on the degree of cancer risk and there are no clear guidelines on risk management for carriers of mutations. Not all genes included on available multi-gene test are necessarily clinically actionable.
• As is the case with high-risk genes, it is possible that the risks associated with moderate-risk genes may not be entirely due to that gene alone, but may be influenced by gene/gene or gene/environment interactions. Therefore, it may be difficult to use a known mutation alone to assign risk for relatives.
• In many cases, the information from testing for moderate penetrance genes does not change risk management compared with that based on family history alone....
• There is an increased likelihood of finding variants of uncertain significance when testing for pathogenic/likely pathogenic variants in multiple genes.
• It is for these and other reasons that multigene testing is ideally offered in the context of professional genetic expertise for pre- and post-test counseling.

National Comprehensive Cancer Network guidelines on genetic/familial high-risk assessment for colorectal cancer (v.2.2019) state that “when more than one gene can explain an inherited cancer syndrome, then multi-gene testing may be more efficient and/or cost-effective than single-gene testing,” and “there is also a role for multi-gene testing in individuals who have tested negative (indeterminate) for a single syndrome, but whose personal or family history remains strongly suggestive of an inherited susceptibility.”20 However, the Network cautioned about the increased likelihood of finding variants of uncertain significance, which increases with the number of genes included in the panel, and that gene panels can include moderate-risk genes that may not be clinically actionable.

U.S. Preventive Services Task Force Recommendations
The U.S. Preventive Services Task Force (2015) has recommended that primary care providers screen women who have family members with breast, ovarian, tubal, or peritoneal cancer with one of several screening tools designed to identify a family history that may be associated with an increased risk for potentially harmful variants in breast cancer and susceptibility genes (BRCA1 or BRCA2).21 Women with positive screening results should receive genetic counseling and if indicated after counseling, BRCA testing (grade B recommendation). The use of genetic cancer susceptibility panels was not specifically mentioned.

Medicare National Coverage
There is no national coverage determination. In the absence of a national coverage determination, coverage decisions are left to the discretion of local Medicare carriers.

Ongoing and Unpublished Clinical Trials
Some currently ongoing and unpublished trials that might influence this review are listed in Table 6.

Table 6. Summary of Key Trials

<table>
<thead>
<tr>
<th>NCT No.</th>
<th>Trial Name</th>
<th>Planned Enrollment</th>
<th>Completion Date</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ongoing</td>
<td>NCT03688204 Clinical Implementation of a Polygenic Risk Score (PRS) for Breast Cancer: Impact on Risk Estimates, Management Recommendations, Clinical Outcomes, and Patient Perception</td>
<td>2,000</td>
<td>Sep 2025</td>
</tr>
<tr>
<td>Unpublished</td>
<td></td>
<td></td>
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</table>

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References


**Documentation for Clinical Review**

- No records required

**Coding**

This Policy relates only to the services or supplies described herein. Benefits may vary according to product design; therefore, contract language should be reviewed before applying the terms of the Policy. Inclusion or exclusion of codes does not constitute or imply member coverage or provider reimbursement.

**IE**

The following services may be considered investigational.

<table>
<thead>
<tr>
<th>Type</th>
<th>Code</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>CPT®</td>
<td>0048U</td>
<td>Oncology (solid organ neoplasia), DNA, targeted sequencing of protein-coding exons of 468 cancer-associated genes, including interrogation for somatic mutations and microsatellite instability, matched with normal specimens, utilizing formalin-fixed paraffin-embedded tumor tissue, report of clinically significant mutation(s)</td>
</tr>
<tr>
<td></td>
<td>0101U</td>
<td>Hereditary colon cancer disorders (e.g., Lynch syndrome, PTEN hamartoma syndrome, Cowden syndrome, familial adenomatosis polyposis); genomic sequence analysis panel utilizing a combination of NGS, Sanger, MLPA and array CGH, with mRNA analytics to resolve variants of unknown significance when indicated [15 genes (sequencing and deletion/duplication), EPCAM and GREM1 (deletion/duplication only)] (Code effective 7/1/2019)</td>
</tr>
<tr>
<td></td>
<td>0102U</td>
<td>Hereditary breast cancer-related disorders (e.g., hereditary breast cancer, hereditary ovarian cancer, hereditary endometrial cancer); genomic sequence analysis panel utilizing a combination of NGS, Sanger, MLPA and array CGH, with MRNA analytics to resolve variants of unknown significance when indicated [17 genes (sequencing and deletion/duplication)] (Code effective 7/1/2019)</td>
</tr>
<tr>
<td></td>
<td>0103U</td>
<td>Hereditary ovarian cancer (e.g., hereditary ovarian cancer, hereditary endometrial cancer); genomic sequence analysis panel utilizing a combination of NGS, Sanger, MLPA and array CGH, with MRNA analytics to resolve variants of unknown significance when indicated [13 genes (sequencing and deletion/duplication)]</td>
</tr>
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</table>

Reproduction without authorization from Blue Shield of California is prohibited
<table>
<thead>
<tr>
<th>Type</th>
<th>Code</th>
<th>Description</th>
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</thead>
<tbody>
<tr>
<td>0104U</td>
<td>Hereditary pan cancer (e.g., hereditary breast and ovarian cancer, hereditary endometrial cancer, hereditary colorectal cancer); genomic sequence analysis panel utilizing a combination of NGS, Sanger, MLPA and array CGH, with MRNA analytics to resolve variants of unknown significance when indicated [32 genes (sequencing and deletion/duplication); EPCAM and GREM1 (deletion/duplication only)]</td>
<td>(Code effective 7/1/2019)</td>
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<tr>
<td>0129U</td>
<td>Hereditary breast cancer-related disorders (e.g., hereditary breast cancer, hereditary ovarian cancer, hereditary endometrial cancer), genomic sequence analysis and deletion/duplication analysis panel (ATM, BRCA1, BRCA2, CDH1, CHEK2, PALB2, PTEN, and TP53)</td>
<td>(Deleted code effective 10/1/2019)</td>
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<tr>
<td>0130U</td>
<td>Hereditary colon cancer disorders (e.g., Lynch syndrome, PTEN hamartoma syndrome, Cowden syndrome, familial adenomatosis polyposis), targeted mRNA sequence analysis panel (APC, CDH1, CHEK2, MLH1, MSH2, MSH6, MUYTH, PMS2, PTEN, and TP53) (List separately in addition to code for primary procedure)</td>
<td>(Code effective 10/1/2019)</td>
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<tr>
<td>0131U</td>
<td>Hereditary breast cancer-related disorders (e.g., hereditary breast cancer, hereditary ovarian cancer, hereditary endometrial cancer), targeted mRNA sequence analysis panel (13 genes)</td>
<td>(Code effective 10/1/2019)</td>
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<tr>
<td>0132U</td>
<td>Hereditary ovarian cancer-related disorders (e.g., hereditary breast cancer, hereditary ovarian cancer, hereditary endometrial cancer), targeted mRNA sequence analysis panel (17 genes)</td>
<td>(List separately in addition to code for primary procedure) (Code effective 10/1/2019)</td>
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<tr>
<td>0133U</td>
<td>Hereditary prostate cancer-related disorders, targeted mRNA sequence analysis panel (11 genes)</td>
<td>(List separately in addition to code for primary procedure) (Code effective 10/1/2019)</td>
</tr>
<tr>
<td>0134U</td>
<td>Hereditary pan cancer (e.g., hereditary breast and ovarian cancer, hereditary endometrial cancer, hereditary colorectal cancer), targeted mRNA sequence analysis panel (18 genes)</td>
<td>(List separately in addition to code for primary procedure) (Code effective 10/1/2019)</td>
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<tr>
<td>0135U</td>
<td>Hereditary gynecological cancer (e.g., hereditary breast and ovarian cancer, hereditary endometrial cancer, hereditary colorectal cancer), targeted mRNA sequence analysis panel (12 genes)</td>
<td>(List separately in addition to code for primary procedure) (Code effective 10/1/2019)</td>
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<tr>
<td>0136U</td>
<td>ATM (ataxia telangiectasia mutated) (e.g., ataxia telangiectasia) mRNA sequence analysis</td>
<td>(List separately in addition to code for primary procedure) (Code effective 10/1/2019)</td>
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<tr>
<td>0137U</td>
<td>PALB2 (partner and localizer of BRCA2) (e.g., breast and pancreatic cancer) mRNA sequence analysis</td>
<td>(List separately in addition to code for primary procedure) (Code effective 10/1/2019)</td>
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<tr>
<td>0138U</td>
<td>BRCA1 (BRCA1, DNA repair associated), BRCA2 (BRCA2, DNA repair associated) (e.g., hereditary breast and ovarian cancer) mRNA sequence analysis</td>
<td>(List separately in addition to code for primary procedure) (Code effective 10/1/2019)</td>
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<tr>
<td>Type</td>
<td>Code</td>
<td>Description</td>
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<tr>
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<td>81120</td>
<td>IDH1 (isocitrate dehydrogenase 1 [NADP+], soluble) (e.g., glioma), common variants (e.g., R132H, R132C)</td>
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<tr>
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<td>81121</td>
<td>IDH2 (isocitrate dehydrogenase 2 [NADP+], mitochondrial) (e.g., glioma), common variants (e.g., R140W, R172M)</td>
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<tr>
<td></td>
<td>81163</td>
<td>BRCA1 (BRCA1, DNA repair associated), BRCA2 (BRCA2, DNA repair associated) (e.g., hereditary breast and ovarian cancer) gene analysis; full sequence analysis (Code effective 1/1/2019)</td>
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<tr>
<td></td>
<td>81164</td>
<td>BRCA1 (BRCA1, DNA repair associated), BRCA2 (BRCA2, DNA repair associated) (e.g., hereditary breast and ovarian cancer) gene analysis; full duplication/deletion analysis (i.e., detection of large gene rearrangements) (Code effective 1/1/2019)</td>
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<tr>
<td></td>
<td>81165</td>
<td>BRCA1 (BRCA1, DNA repair associated) (e.g., hereditary breast and ovarian cancer) gene analysis; full sequence analysis (Code effective 1/1/2019)</td>
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<td>81166</td>
<td>BRCA1 (BRCA1, DNA repair associated) (e.g., hereditary breast and ovarian cancer) gene analysis; full duplication/deletion analysis (i.e., detection of large gene rearrangements) (Code effective 1/1/2019)</td>
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<td>81167</td>
<td>BRCA2 (BRCA2, DNA repair associated) (e.g., hereditary breast and ovarian cancer) gene analysis; full duplication/deletion analysis (i.e., detection of large gene rearrangements) (Code effective 1/1/2019)</td>
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<td>81201</td>
<td>APC (adenomatous polyposis coli) (e.g., familial adenomatosis polyposis [FAP], attenuated FAP) gene analysis; full gene sequence</td>
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<tr>
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<td>81203</td>
<td>APC (adenomatous polyposis coli) (e.g., familial adenomatosis polyposis [FAP], attenuated FAP) gene analysis; duplication/deletion variants</td>
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<td></td>
<td>81211</td>
<td>BRCA1, BRCA2 (breast cancer 1 and 2) (e.g., hereditary breast and ovarian cancer) gene analysis; full sequence analysis and common duplication/deletion variants in BRCA1 (i.e., exon 13 del 3.835kb, exon 13 dup 6kb, exon 14-20 del 26kb, exon 22 del 510bp, exon 8-9 del 7.1kb) (Deleted code effective 1/1/2019)</td>
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<tr>
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<td>BRCA1, BRCA2 (breast cancer 1 and 2) (e.g., hereditary breast and ovarian cancer) gene analysis; uncommon duplication/deletion variants (Deleted code effective 1/1/2019)</td>
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<tr>
<td></td>
<td>81228</td>
<td>Cytogenomic constitutional (genome-wide) microarray analysis; interrogation of genomic regions for copy number variants (e.g., bacterial artificial chromosome [BAC] or oligo-based comparative genomic hybridization [CGH] microarray analysis)</td>
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<tr>
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<td>81292</td>
<td>MLH1 (mutL homolog 1, colon cancer, nonpolyposis type 2) (e.g., hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; full sequence analysis</td>
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<td>81294</td>
<td>MLH1 (mutL homolog 1, colon cancer, nonpolyposis type 2) (e.g., hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; duplication/deletion variants</td>
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<tr>
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<td>81295</td>
<td>MSH2 (mutS homolog 2, colon cancer, nonpolyposis type 1) (e.g., hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; full sequence analysis</td>
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<tr>
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<td>81297</td>
<td>MSH2 (mutS homolog 2, colon cancer, nonpolyposis type 1) (e.g., hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; duplication/deletion variants</td>
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<tr>
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<td>81298</td>
<td>MSH6 (mutS homolog 6 [E. coli]) (e.g., hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; full sequence analysis</td>
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<td>Type</td>
<td>Code</td>
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<tr>
<td></td>
<td>81300</td>
<td>MSH6 (mutS homolog 6 [E. coli]) (e.g., hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; duplication/deletion variants</td>
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<td>81317</td>
<td>PMS2 (postmeiotic segregation increased 2 [S. cerevisiae]) (e.g., hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; full sequence analysis</td>
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<tr>
<td></td>
<td>81319</td>
<td>PMS2 (postmeiotic segregation increased 2 [S. cerevisiae]) (e.g., hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; duplication/deletion variants</td>
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<tr>
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<td>81321</td>
<td>PTEN (phosphatase and tensin homolog) (e.g., Cowden syndrome, PTEN hamartoma tumor syndrome) gene analysis; full sequence analysis</td>
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<td>81323</td>
<td>PTEN (phosphatase and tensin homolog) (e.g., Cowden syndrome, PTEN hamartoma tumor syndrome) gene analysis; duplication/deletion variant</td>
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<td></td>
<td>81404</td>
<td>Molecular pathology procedure level 5</td>
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<td></td>
<td>81408</td>
<td>Molecular pathology procedure level 9</td>
</tr>
<tr>
<td></td>
<td>81432</td>
<td>Hereditary breast cancer-related disorders (e.g., hereditary breast cancer, hereditary ovarian cancer, hereditary endometrial cancer); genomic sequence analysis panel, must include sequencing of at least 10 genes, always including BRCA1, BRCA2, CDH1, MLH1, MSH2, MSH6, PALB2, PTEN, STK11, and TP53</td>
</tr>
<tr>
<td></td>
<td>81433</td>
<td>Hereditary breast cancer-related disorders (e.g., hereditary breast cancer, hereditary ovarian cancer, hereditary endometrial cancer); duplication/deletion analysis panel, must include analyses for BRCA1, BRCA2, MLH1, MSH2, and STK11</td>
</tr>
<tr>
<td></td>
<td>81435</td>
<td>Hereditary colon cancer disorders (e.g., Lynch syndrome, PTEN hamartoma syndrome, Cowden syndrome, familial adenomatosis polyposis); genomic sequence analysis panel, must include sequencing of at least 10 genes, including APC, BMPR1A, CDH1, MLH1, MSH2, MSH6, MUTYH, PTEN, SMAD4, and STK11</td>
</tr>
<tr>
<td></td>
<td>81436</td>
<td>Hereditary colon cancer disorders (e.g., Lynch syndrome, PTEN hamartoma syndrome, Cowden syndrome, familial adenomatosis polyposis); duplication/deletion analysis panel, must include analysis of at least 5 genes, including MLH1, MSH2, EPCAM, SMAD4, and STK11</td>
</tr>
<tr>
<td></td>
<td>81437</td>
<td>Hereditary neuroendocrine tumor disorders (e.g., medullary thyroid carcinoma, parathyroid carcinoma, malignant pheochromocytoma or paraganglioma); genomic sequence analysis panel, must include sequencing of at least 6 genes, including MAX, SDHB, SDHC, SDHD, TMEM127, and VHL</td>
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<tr>
<td></td>
<td>81438</td>
<td>Hereditary neuroendocrine tumor disorders (e.g., medullary thyroid carcinoma, parathyroid carcinoma, malignant pheochromocytoma or paraganglioma); duplication/deletion analysis panel, must include analyses for SDHB, SDHC, SDHD, and VHL</td>
</tr>
<tr>
<td></td>
<td>81445</td>
<td>Targeted genomic sequence analysis panel, solid organ neoplasm, DNA analysis, and RNA analysis when performed, 5-50 genes (e.g., ALK, BRAF, CDKN2A, EGFR, ERBB2, KIT, KRAS, NRAS, MET, PDGFRA, PDGFRB, PGR, PIK3CA, PTEN, RET), interrogation for sequence variants and copy number variants or rearrangements, if performed</td>
</tr>
</tbody>
</table>
### Type | Code | Description
--- | --- | ---
 | 81450 | Targeted genomic sequence analysis panel, hematolymphoid neoplasm or disorder, DNA analysis, and RNA analysis when performed, 5-50 genes (e.g., BRAF, CEBPA, DNMT3A, EZH2, FLT3, IDH1, IDH2, JAK2, KRAS, KIT, MLL, NRAS, NPM1, NOTCH1), interrogation for sequence variants, and copy number variants or rearrangements, or isoform expression or mRNA expression levels, if performed.
 | 81455 | Targeted genomic sequence analysis panel, solid organ or hematolymphoid neoplasm, DNA analysis, and RNA analysis when performed, 51 or greater genes (e.g., ALK, BRAF, CDKN2A, CEBPA, DNMT3A, EGFR, ERBB2, EZH2, FLT3, IDH1, IDH2, JAK2, KIT, KRAS, MLL, NPM1, NRAS, MET, NOTCH1, PDGFA, PDGFRB, PGR, PIK3CA, PTEN, RET), interrogation for sequence variants and copy number variants or rearrangements, if performed.
 | 81479 | Unlisted molecular pathology procedure.

### HCPCS
None

### ICD-10 Procedure
None

### Policy History

This section provides a chronological history of the activities, updates and changes that have occurred with this Medical Policy.

<table>
<thead>
<tr>
<th>Effective Date</th>
<th>Action</th>
<th>Reason</th>
</tr>
</thead>
<tbody>
<tr>
<td>09/27/2013</td>
<td>BCBSA Medical Policy adoption</td>
<td>Medical Policy Committee</td>
</tr>
<tr>
<td>01/30/2015</td>
<td>Coding update</td>
<td>Administrative Review</td>
</tr>
<tr>
<td>06/30/2015</td>
<td>Coding update</td>
<td>Administrative Review</td>
</tr>
<tr>
<td>02/01/2016</td>
<td>Coding update</td>
<td>Administrative Review</td>
</tr>
<tr>
<td>08/01/2016</td>
<td>Policy title change from Genetic Cancer Susceptibility Panels Using Next Generation Sequencing Policy revision without position change</td>
<td>Medical Policy Committee</td>
</tr>
<tr>
<td>09/01/2017</td>
<td>Policy revision without position change</td>
<td>Medical Policy Committee</td>
</tr>
<tr>
<td>12/01/2017</td>
<td>Policy revision without position change</td>
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<tr>
<td>02/01/2018</td>
<td>Coding update</td>
<td>Administrative Review</td>
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<tr>
<td>08/01/2018</td>
<td>Coding update</td>
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<tr>
<td>12/01/2018</td>
<td>Policy revision without position change Coding update</td>
<td>Medical Policy Committee</td>
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<tr>
<td>01/01/2019</td>
<td>Policy statement clarification Coding update</td>
<td>Administrative Review</td>
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<tr>
<td>07/01/2019</td>
<td>Coding update</td>
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<td>11/01/2019</td>
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<tr>
<td>12/01/2019</td>
<td>Policy revision without position change</td>
<td>Medical Policy Committee</td>
</tr>
</tbody>
</table>

### Definitions of Decision Determinations

**Medically Necessary:** A treatment, procedure, or drug is medically necessary only when it has been established as safe and effective for the particular symptoms or diagnosis, is not investigational or experimental, is not being provided primarily for the convenience of the patient or the provider, and is provided at the most appropriate level to treat the condition.
Investigational/Experimental: A treatment, procedure, or drug is investigational when it has not been recognized as safe and effective for use in treating the particular condition in accordance with generally accepted professional medical standards. This includes services where approval by the federal or state governmental is required prior to use, but has not yet been granted.

Split Evaluation: Blue Shield of California/Blue Shield of California Life & Health Insurance Company (Blue Shield) policy review can result in a split evaluation, where a treatment, procedure, or drug will be considered to be investigational for certain indications or conditions, but will be deemed safe and effective for other indications or conditions, and therefore potentially medically necessary in those instances.

Prior Authorization Requirements (as applicable to your plan)

Within five days before the actual date of service, the provider must confirm with Blue Shield that the member’s health plan coverage is still in effect. Blue Shield reserves the right to revoke an authorization prior to services being rendered based on cancellation of the member’s eligibility. Final determination of benefits will be made after review of the claim for limitations or exclusions.

Questions regarding the applicability of this policy should be directed to the Prior Authorization Department. Please call (800) 541-6652 or visit the provider portal at www.blueshieldca.com/provider.

Disclaimer: This medical policy is a guide in evaluating the medical necessity of a particular service or treatment. Blue Shield of California may consider published peer-reviewed scientific literature, national guidelines, and local standards of practice in developing its medical policy. Federal and state law, as well as contract language, including definitions and specific contract provisions/exclusions, take precedence over medical policy and must be considered first in determining covered services. Member contracts may differ in their benefits. Blue Shield reserves the right to review and update policies as appropriate.